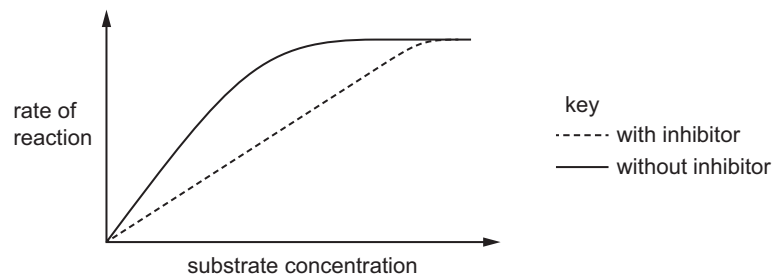


12 The graph shows the effect of substrate concentration on an enzyme-catalysed reaction with and without a competitive inhibitor.



What is the effect of the competitive inhibitor on V_{max} and K_m ?

- A** V_{max} decreases and K_m decreases.
- B** V_{max} stays the same and K_m decreases.
- C** V_{max} stays the same and K_m increases.
- D** V_{max} decreases and K_m stays the same.

14 How is the Michaelis–Menten constant (K_m) used?

- A** to determine the number of collisions between the enzyme and substrate
- B** to compare the affinity of enzymes for their substrates
- C** to find the maximum velocity of an enzyme (V_{max})
- D** to find the optimum rate of reaction

15 Some students investigated the effect of pH on the rate of starch digestion by amylase at 20 °C.

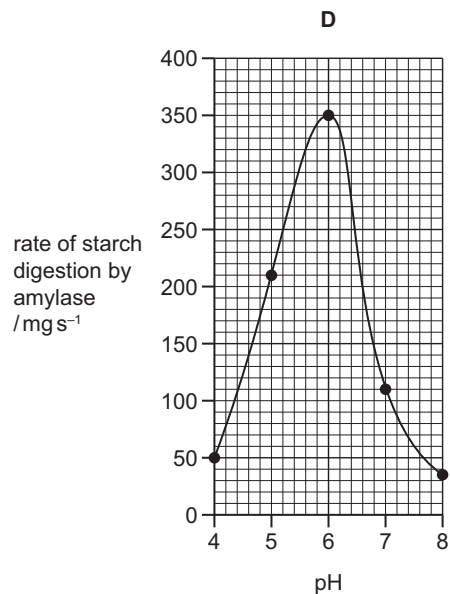
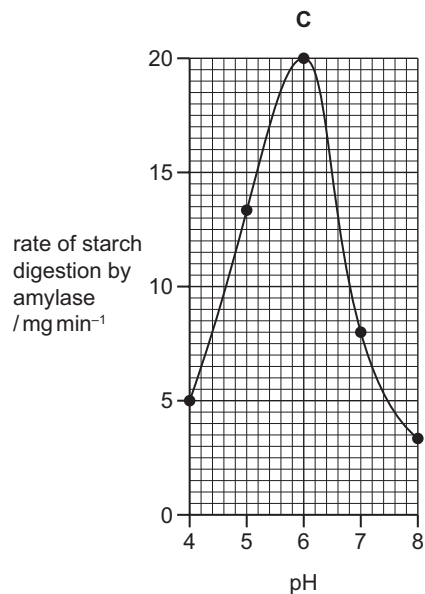
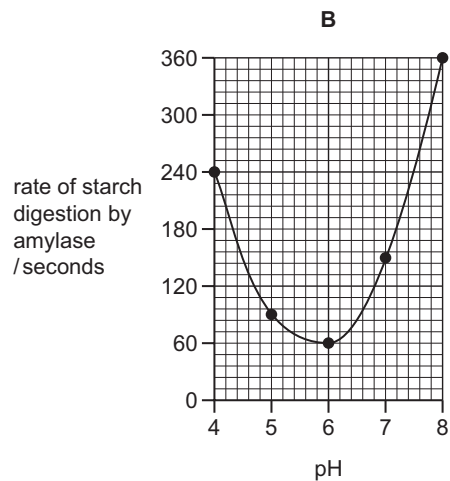
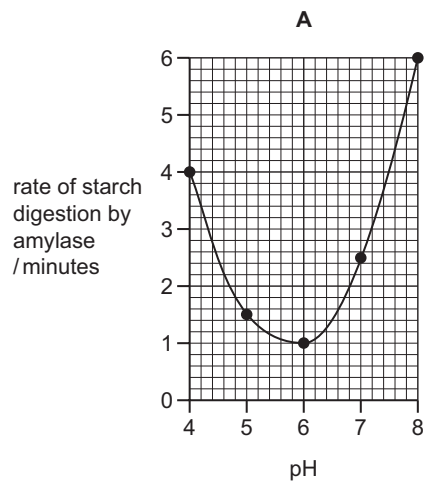
- They added 2 cm³ of a 1% amylase solution and 1 cm³ of pH 4 buffer to a test-tube.
- They added 2 cm³ of a starch solution, containing 0.02 g of starch, to the test-tube.
- They measured the time taken for the starch to be digested (to disappear).

The students then repeated these steps with pH 5, pH 6, pH 7 and pH 8 buffers.

The table shows the results.

pH	time taken for starch to be digested / minutes
4	4.0
5	1.5
6	1.0
7	2.5
8	6.0

Which graph is correct for these data?

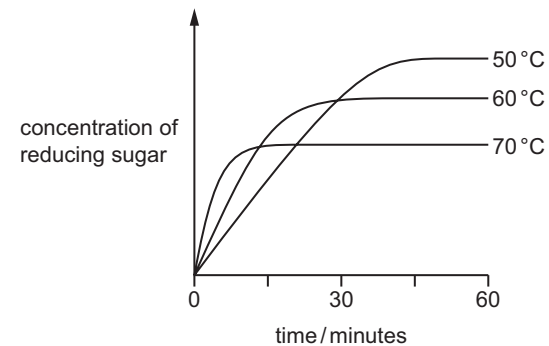


16 Which statements are correct for a non-competitive inhibitor of enzyme action?

- 1 Increasing the concentration of the enzyme's substrate will reduce its effect.
- 2 It reduces the activation energy required for a reaction to take place.
- 3 It reduces the maximum rate of reaction.

A 1 and 3 **B** 1 only **C** 2 and 3 **D** 3 only

16 The graph shows the results of an investigation into the effect of amylase on starch at three different temperatures.



Which statements are correct conclusions using these results?

- 1 The optimum temperature is 50 °C.
- 2 The initial rate of reaction is highest at 70 °C.
- 3 The higher the temperature the more quickly the enzyme denatures.

A 1, 2 and 3 **B** 1 and 2 only **C** 1 and 3 only **D** 2 and 3 only

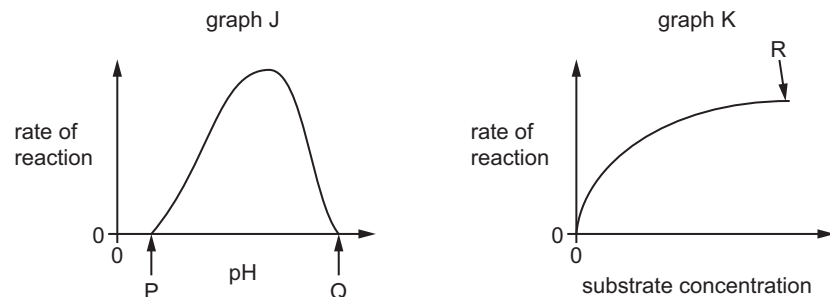
17 The rate of enzyme-catalysed reactions in human cells is regulated.

What may be involved in this regulation?

- 1 a change in enzyme concentration
- 2 a change in substrate concentration
- 3 inhibition by the final product of the reaction

A 1, 2 and 3 **B** 1 and 2 only **C** 1 and 3 only **D** 2 and 3 only

15 Graph J shows the effect of pH on the rate of an enzyme reaction. Graph K shows the effect of substrate concentration on the rate of an enzyme reaction.



A student wrote four statements about the graphs.

- 1 In graph J, the enzyme is denatured at point Q.
- 2 In graph J, the enzyme is inactive at point P.
- 3 In graph K, the rate of reaction is directly proportional to the substrate concentration.
- 4 In graph K, $V_{\max}$ is reached at point R.

Which statements are correct?

- A** 1, 2 and 3 **B** 1, 2 and 4 **C** 1, 3 and 4 **D** 2, 3 and 4

- 1 The bacterium *Streptococcus pneumoniae* is commonly found in the throat. *S. pneumoniae* produces hydrogen peroxide as it grows. A sample can be taken from a patient's throat and tested to measure the concentration of hydrogen peroxide. This can be used as a measure of the growth of the bacteria.

You will determine the growth of bacteria by measuring the concentration of hydrogen peroxide in a solution that represents a sample taken from a patient. You will do this by measuring how long it takes for a sample of hydrogen peroxide to cause a colour change in a reaction mixture. The faster the mixture changes to a blue-black colour, the higher the concentration of hydrogen peroxide, and the greater the growth of bacteria.

You will use a range of known concentrations of hydrogen peroxide to estimate the concentration of hydrogen peroxide in a sample.

You are provided with the materials shown in Table 1.1.

Table 1.1

labelled	contents	hazard	volume / cm ³
R1	dilute sulfuric acid	irritant	100
R2	starch solution	low	10
R3	potassium iodide solution	low	10
R4	sodium thiosulfate solution	low	10
H	2.0% hydrogen peroxide solution	irritant	25
U	solution representing patient sample	irritant	10
W	distilled water	low	100

If any solution comes into contact with your skin, wash off immediately with cold water.

It is recommended that you wear suitable eye protection.

You will need to carry out a **serial** dilution of the 2.0% hydrogen peroxide solution, **H**, to reduce the concentration by **half** between each successive dilution.

You will need to prepare **four** concentrations of hydrogen peroxide solution in addition to the 2.0% hydrogen peroxide solution, **H**.

After the serial dilution is completed, you will need to have 10 cm³ of each concentration available to use.

- (a) (i)** Complete Fig. 1.1 to show how you will prepare your serial dilution.

Each beaker should have:

- a labelled arrow to show the volume of hydrogen peroxide solution transferred
- a labelled arrow to show the volume of distilled water, **W**, added
- a label under the beaker to show the concentration of the hydrogen peroxide solution.

Name:

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0 cm³ of **W**